Structure and Absolute Configuration of Solanapyrone D: A New Clue to the

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The structure and absolute configuration of solanapyrone D (2) offers further indirect evidence that solanapyrone A (1) and D (2) are biosynthesized *via* intramolecular Diels–Alder reactions.

Involvement of biological Diels–Alder reactions has been proposed in the biosynthesis of a variety of secondary metabolites.¹ Among the candidates for such a biogenetic path are the alkaloid presecamines,² the terpenoid heliocides,³ and the polyketide cytochalacins.⁴ However, these compounds have rarely been isolated together with their optically active regioisomer or diastereoisomer. Here we report the concurrence and isolation of an optically active diastereoisomer of solanapyrone A (1) which would result from *endo* cycloaddition in a biological Diels–Alder reaction.

Occurrence of Biological Diels–Alder Reactions



Figure 1

During the biosynthetic study of (1), compound (2), solanapyrone D, was detected and isolated as a minor component. This material was optically active $\{[\alpha]_D^{24} - 125.2^{\circ} (c \ 0.8, CHCl_3)\}$, and was considered as a diastereoisomer of (1) from its molecular formula $(C_{18}H_{22}O_4)$ and various spectral data. Eventually, the structure was identified as (2), depicted in Figure 1, which had been derived from a major Diels-Alder adduct in the reaction of triene (3a) (Figure 1).⁵

Determination of the absolute configuration of (2) also provides valuable information about the enzymatic reaction. This was undertaken as follows (Figure 2). A 4:1 mixture of (1) and (2) was treated with OsO_4 at -20 °C to give products (4a) and (4b), respectively. In this oxidation, the reagent attacks stereoselectively from less hindered sides, *i.e.* the convex face in (1) and the side opposite to the axial methyl group in (2). The products (4a) and (4b) are separated after



Figure 2





conversion to di-*p*-methoxybenzoates (**5a**) and (**5b**).[†] Based on the ¹H n.m.r. spectroscopy coupling constants, the relationship between 3-H and 4-H was assigned as equatorialaxial in (**5a**), and as axial-equatorial in (**5b**). The negative split in the Cotton effect [λ_{max} . (EtOH) ($\Delta \varepsilon$): 268 (-15.5), 248 (+12.3) nm (c 0.032 mM)] was observed in the c.d. spectrum of (**5b**). From this result, the absolute configuration of (**2**) was assigned as depicted in Figure 1. Interaction between the *p*-methoxybenzoyl group and the pyrone moiety can be ignored because of the distance between these groups and the different location of their u.v. maxima.

The stereochemistry at C-1 and C-10 in (2) is the same as that of (1) whereas the configurations at C-2 and C-5 in (2) are opposite to those of (1).⁶ These data allow speculation that the

postulated enzyme which catalyses the Diels-Alder reaction recognizes the dienophile portion and terminal methyl group, and accepts two similar transition states (A) and (B) of (3), which can be interconverted by rotation about the C-5 to C-6 bond (Scheme 1). Thus, transition state (A) (exo) gives (1), and (B) (endo) gives (2). Consideration of Dreiding models indicates that the stability of the two transition states would be similar. In fact, this is reflected in the product ratio [(1a)/(2a)]= 1/2] in the synthesis of (1).⁵ In more than 30 decalin polyketides,7 solanapyrones represent the only case in which the triene precursor [e.g., (3)] does not possess any chiral centre. Possibly this accounts for the biological production of two diastereoisomers. Interestingly, kuwanones X (6) and Y (7), a pair of diastereoisomers possibly arising from exo and endo intermolecular cycloaddition, respectively, have the same stereochemical relationship as (1) and (2) (Figure 3).⁸

In conclusion, stereochemical information and isolation of a pair of possible adducts strongly suggest that (1) and (2) are biosynthesized via an intramolecular Diels-Alder reaction. The timing of oxidation of the C_1 unit at the pyrone ring is presently being investigated prior to incorporation experiments with trienes.

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[†] Spectroscopic data. ¹H n.m.r. (CDCl₃). (**5a**): δ 8.08 (2 H, d, J 8.8 Hz), 7.83 (2 H, d, J 8.9 Hz), 7.00 (2 H, d, J 8.8 Hz), 6.81 (2 H, d, J 8.9 Hz), 5.38 (1 H, dd, J 2.8, 3.0 Hz, 4-H), 5.19 (1 H, dd, J 2.8, 11.0 Hz, 3-H), 3.90 (3 H, s), 3.81 (3 H, s). (**5b**) δ 8.05 (2 H, d, J 8.8 Hz), 7.74 (2 H, d, J 8.9 Hz), 7.01 (2 H, d, J 8.8 Hz), 6.82 (2 H, d, J 8.9 Hz), 5.45 (1 H, dd, J 2.6, 2.8 Hz, 3-H), 5.22 (1 H, dd, J 2.6, 11.5 Hz, 4-H), 3.91 (3 H, s), 3.81 (3 H, s).